

# „Donor suitability and donation process“

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# Donor suitability and donation process

1794

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## BRIEF REPORT

### INADVERTENT TRANSMISSION OF A DONOR'S ACUTE MYELOID LEUKEMIA IN BONE MARROW TRANSPLANTATION FOR CHRONIC MYELOCYTIC LEUKEMIA

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layered with 0.25 ml of medium containing various concentrations of serum samples from the patient or a control. After incubation for 12 days at 37°C, 1 ml of 2.5 percent glutaraldehyde was injected into the well to detach and fix the agar layer. Cell aggregates containing more than 50 granulocytic and monocytic cells were scored as granulomonocytic colonies.

#### Analysis of Surface-Antigen Patterns

The expression of different surface antigens of bone marrow cells was measured by direct immunofluorescence and flow cytometry on a fluorescence-activated cell sorter (Becton Dickinson, Mountain View, Calif.) or by immunohistologic examination of cryostat sections of bone marrow. In the analysis of surface markers, the following monoclonal antibodies were applied: M59 (CD33, Coulter, Hialeah, Fla.), M57 (CD13, Coulter), VIM-2 (Dr. Walter Knapp, University of Vienna, Austria), BMA 0200 (CD15, Behringwerke, Marburg, Federal Republic of Germany), anti-HLA-DR (Becton

# Donor suitability and donation process

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## Mini review

### **Transmission of donor illness by stem cell transplantation: should screening be different in older donors?**

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# Donor suitability and donation process

**Table 1** Transmission of donor diseases to recipients

<i>Transmission of</i>	<i>Donor disease</i>
<i>Infectious diseases</i>	
Viruses	HIV <sup>1,2</sup> Hepatitis B and C <sup>1,2</sup> HTLV-1 <sup>1,3</sup> CMV <sup>1,4-16</sup> EBV Parvovirus B19 <sup>1,7</sup> West Nile <sup>1,8,19</sup>
Bacteria	Contaminants <sup>a</sup> Brucellosis <sup>20</sup>
Parasites	Toxoplasmosis <sup>21,22</sup> Malaria <sup>23-26</sup> Leishmania <sup>1,2</sup> Babesia <sup>1,2</sup>
Fungi	Candida, Aspergillus
Prions	Creutzfeldt Jakob disease <sup>b</sup>

<sup>a</sup>Contamination rates of one in 3000 units in platelet concentrates. No information available on stem cell grafts.

<sup>b</sup>No known cases in HCT and blood transfusions.

<sup>c</sup>Not reported in HCT.

# Donor suitability and donation process

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**Table 1** Transmission of donor diseases to recipients

<i>Transmission of</i>	<i>Donor disease</i>
<i>Acquired disorders</i>	
Autoimmune diseases	Myasthenia gravis <sup>28</sup> Atopy <sup>29</sup> SLE specific autantibodies <sup>30</sup> Thyreotoxicosis <sup>31-33</sup> Diabetes mellitus type I <sup>34</sup> Sarcoidosis <sup>35</sup> Coeliac disease <sup>36</sup> Autoimmune thrombocytopenia <sup>37</sup>
Hematological malignancies	AML <sup>38</sup> CML <sup>39</sup> T-cell lymphoma <sup>55</sup>
Nonhematological malignancies <sup>c</sup>	Small-cell lung cancer from renal transplantation <sup>40</sup> Glioblastoma multiforme from liver transplantation <sup>41</sup>

<sup>a</sup>Contamination rates of one in 3000 units in platelet concentrates. No information available on stem cell grafts.

<sup>b</sup>No known cases in HCT and blood transfusions.

<sup>c</sup>Not reported in HCT.

# Donor suitability and donation process

**Table 2** Recommendations for donor work-up

<i>Test</i>	<i>WMDA<sup>61</sup></i>	<i>NMDP 18th Edition Standards/ September 2002</i>	<i>JACIE 2 edition/June 2003</i>	<i>Proposal</i>
Medical history	Yes	Infectious (including risk for infections), pregnancy, blood donation history	Vaccination, travel and blood transfusion history, questions to identify persons at high risk for transmittable infections	Familiar, travel, vaccination, smoking, blood transfusion and donation, infectious, pregnancy, allergy, autoimmune, vaccination and tumor history
Physical examination	Yes	Yes	Yes	Particular attention to cardiovascular, bleeding and malignant diseases
ECG	Yes	Yes	Not specified	Yes
Chest X-ray	Yes	Yes	Not specified	Yes
Blood counts	Full blood count and differential with blood film	Complete blood count	Not specified	Complete blood count, manual differential on blood film
Blood group	Yes with antibody screen	ABO group and Rh type	ABO group and Rh type and appropriate red cell compatibility with the recipient	ABO group and Rh type and appropriate red cell compatibility with the recipient
Coagulation screen	Yes	No	No	<b>PT, PTT</b> , fibrinogen
HLA-typing	Yes	Yes	Yes	HLA typing HLA-A, -B, -C, DRB1, HLA-DQ
Biochemical profile	Urea, electrolytes, creatinine, liver function tests, blood sugar	Urinalysis, electrolytes, blood urea nitrogen or creatinine, bilirubin, serum protein plus albumin or serum protein electrophoresis, hemoglobin S for PBSC donors only	Not specified in detail	Urea, electrolytes, creatinine, liver function tests (bili, ASAT, ALAT, AP, $\gamma$ -GT), LDH, blood sugar, protein electrophoresis, urine analysis
Infectious disease markers (within 30 days prior to collection)	Syphilis, hepatitis B surface antigen, HIV antigen and antibodies, hepatitis B core antigen, hepatitis C, HTLV-1, herpes simplex virus, CMV, <i>Vaccinia zoster</i> virus, and EBV	HIV type 1 and 2, HBV, HCV, HTLV type 1 and 2, <i>Treponema pallidum</i> (syphilis), CMV	HIV type 1 and 2, HBV, HCV, HTLV type I and II, <i>Treponema pallidum</i> and CMV	HIV type 1 and 2, HBV (HBs Ag, HBsAb, HBcAb) HCV (HCVAb), HTLV type I and II, <i>Treponema pallidum</i> , CMV (IgG, IgM), Toxoplasmosis (IgG, IgM) and EBV
Pregnancy test (when indicated)	Yes	Yes	Yes	Gynecological visit including pregnancy test and physical breast examination
Malignant diseases in donors > 55 years of age (related HCT)				PSA in males, physical, occult blood in the stool. Bone marrow aspiration if medical history or tests are abnormal. CT chest scan in case of a long smoking history
Congenital disorders				Testing for congenital disorders of planned recipient within family donors; testing for congenital disorders of the family

# Donor suitability and donation process

**Table 3** Donor selection in case of donors with abnormal findings: (a) family donors, (b) unrelated donors and (c) unrelated cord blood

<i>Abnormal finding on donors</i>	<i>Absolute contraindications</i>	<i>Relative contraindications</i>	<i>Specific consideration</i>
<i>(a) Family donors<sup>a</sup></i>			
Infectious	HIV, HTLV-1,	Lues, HBV, HCV, malaria after 3 years; parvovirus B19	EBV, CMV, toxoplasmosis
Congenital	Thalassemia major; combined immune deficiency. Congenital disease with severely reduced life expectancy	Thalassemia minor; M Gaucher	Targeted cord blood should be tested for congenital disease of planned recipient
Malignancies	Every malignancies except <i>in situ</i> cancer	Skin cancer removed <i>in toto</i>	
Pregnancy		Marrow donation and G-CSF-stimulated apheresis, unstimulated apheresis	
<i>(b) Unrelated donors<sup>a</sup></i>			
Infectious	As for blood donation	Parvovirus B19, if known after collection: Gram-positive graft infection	EBV, CMV, toxoplasmosis
Congenital	As for blood donation		
Malignancies	Every malignancies except <i>in situ</i> cancer	Skin cancer removed <i>in toto</i>	
Pregnancy	Any donation		
<i>(c) Unrelated cord blood<sup>a</sup></i>			
Infectious	As for blood donation Gram-positive, Gram-negative contamination		EBV, CMV, toxoplasmosis
Congenital	As for blood donation; exclude, if congenital diseases known in family		
Malignancies	Any, in child		

<sup>a</sup>No enough information is available for West Nile virus. Contamination of stem cell graft with epidermal bacteria might be a relative contraindication.

# Donor suitability and donation process

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## ORIGINAL ARTICLE

### **The impact of the age of HLA-identical siblings on mobilization and collection of PBSCs for allogeneic hematopoietic cell transplantation**

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# Donor suitability and donation process

**Table 1** Characteristics of patients and their HLA-identical donors  
(n = 167)

<i>Patients</i>	
Median age (years)	48 (18-74)
Gender	No. (%)
Male	95 (57)
Female	72 (43)
<i>Diagnosis</i>	
Acute leukemia	67 (40)
Chronic leukemia	37 (22)
MM	24 (14)
MDS	13 (8)
Others	26 (16)
<i>Conditioning regimens</i>	
RIC	88 (52.7)
Conventional	79 (47.3)
<i>Donors</i>	
Median age (years)	47 (18-74)
< 50 years	100 (59.9)
≥50-59 years	33 (19.8)
≥60 years	34 (20.3)
<i>Gender</i>	
Male	85 (51)
Female	82 (49)
<i>Patient/donor gender</i>	
Male/male	51 (30.5)
Male/female	44 (26.3)
Female/female	38 (22.8)
Female/male	34 (20.4)
<i>History of hypertension in the donor</i>	
No	75 (45)
Yes	92 (55)
<i>History of drug intake in the donor</i>	
No	111 (66.5)
Yes	56 (33.5)

Abbreviations: MDS = myelodysplastic syndrome; MM = multiple myeloma; RIC = reduced intensity conditioning.

# Donor suitability and donation process

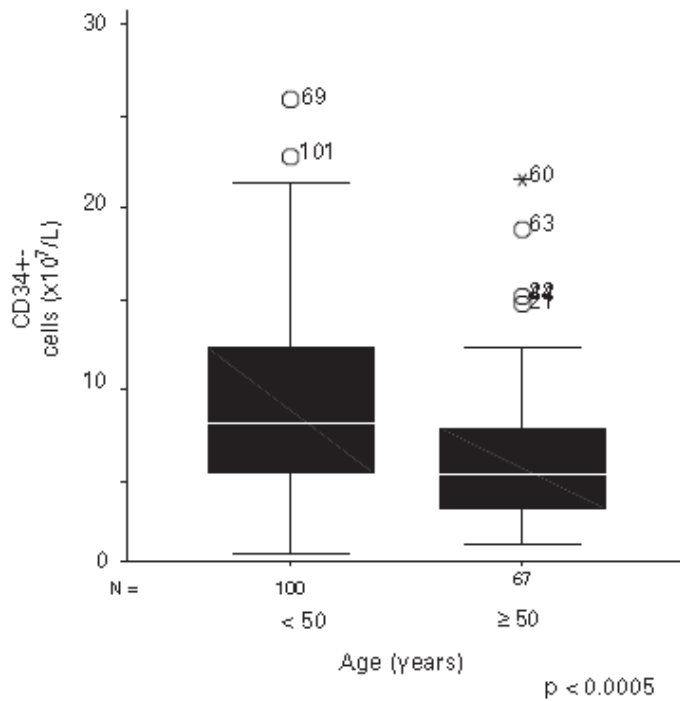


Figure 1 Age of the donor and peripheral blood CD34 + cell count before apheresis.

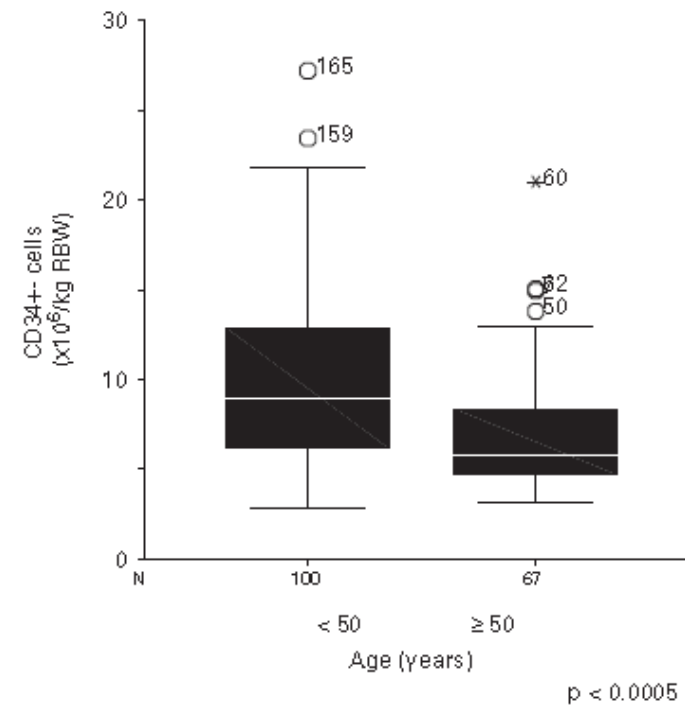


Figure 2 Age of the donors and CD34 + cell counts in the transplants.

# Donor suitability and donation process

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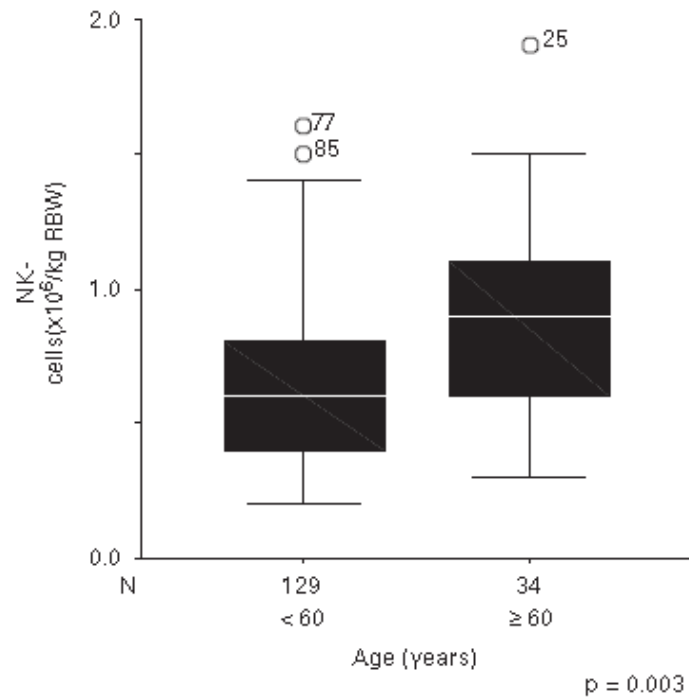


Figure 3 Age of the donor and NK-cell counts in the transplant.

# Donor suitability and donation process

**Table 2** Impact of donors' age on mobilization and harvest of HPC (n = 167)

Parameter	< 60 years vs ≥60 years		< 50 years vs ≥50 years	
	Uni (P-value)	Multi (P-value)	Uni (P-value)	Multi (P-value)
No. of apheresis sessions, median 1 (range 1-3)	0.07	NS	NS	
Filgrastim dose required for mobilization	NS		NS	
Peripheral CD34+ cells before apheresis, median 6.3 (range 1-25.2) × 10 <sup>7</sup> /L	0.001	NS	<0.0005	0.01
<i>Hb, median (range) g per 100 mL</i>				
Before mobilization, 14.3 (11-17.6)	NS		NS	
Before apheresis, 14 (9.5-16.8)	NS		NS	
After apheresis, 13 (8.1-15.8)	NS		NS	
<i>WBC, median (range) × 10<sup>9</sup>/L</i>				
Before mobilization, 6.7 (2.8-14.5)	NS		NS	
Before apheresis, 50.4 (22.5-94.5)	NS		0.04	0.08
After apheresis, 40.9 (10.3-82.8)	NS		0.009	NS
<i>Plts, median (range) × 10<sup>9</sup>/L</i>				
Before mobilization, 246 (127-467)	NS		NS	
Before apheresis, 225 (85-459)	NS		0.09	NS
After apheresis, 94 (22-275)	NS		0.02	NS
CTC toxicity ≥grade II	NS		NS	
<i>Graft, median (range)</i>				
Vol, 295 (83-1499) ml	NS		NS	
CD34+ cells, 7.2 (2.8-33) × 10 <sup>6</sup> /kg	0.003	0.006	<0.0005	NS
CD3+ cells, 3.8 (0.8-21.8) × 10 <sup>6</sup> /kg	NS		NS	
NK cells, 0.7 (0.2-1.9) × 10 <sup>6</sup> /kg	0.003	0.001	0.01	0.07
<i>Non-hematological CTC toxicity ≥ grade II</i>				
AP	0.005	0.07	0.04	NS
PPT	NS		NS	
Potassium	NS		NS	

Abbreviations: AP alkaline phosphatase; CTC the National Cancer Institute Common Toxicity Criteria; Multi multivariate analyses; NK cells natural killer cells; PTT partial thromboplastin time; Uni univariate analyses.

# Donor suitability and donation process

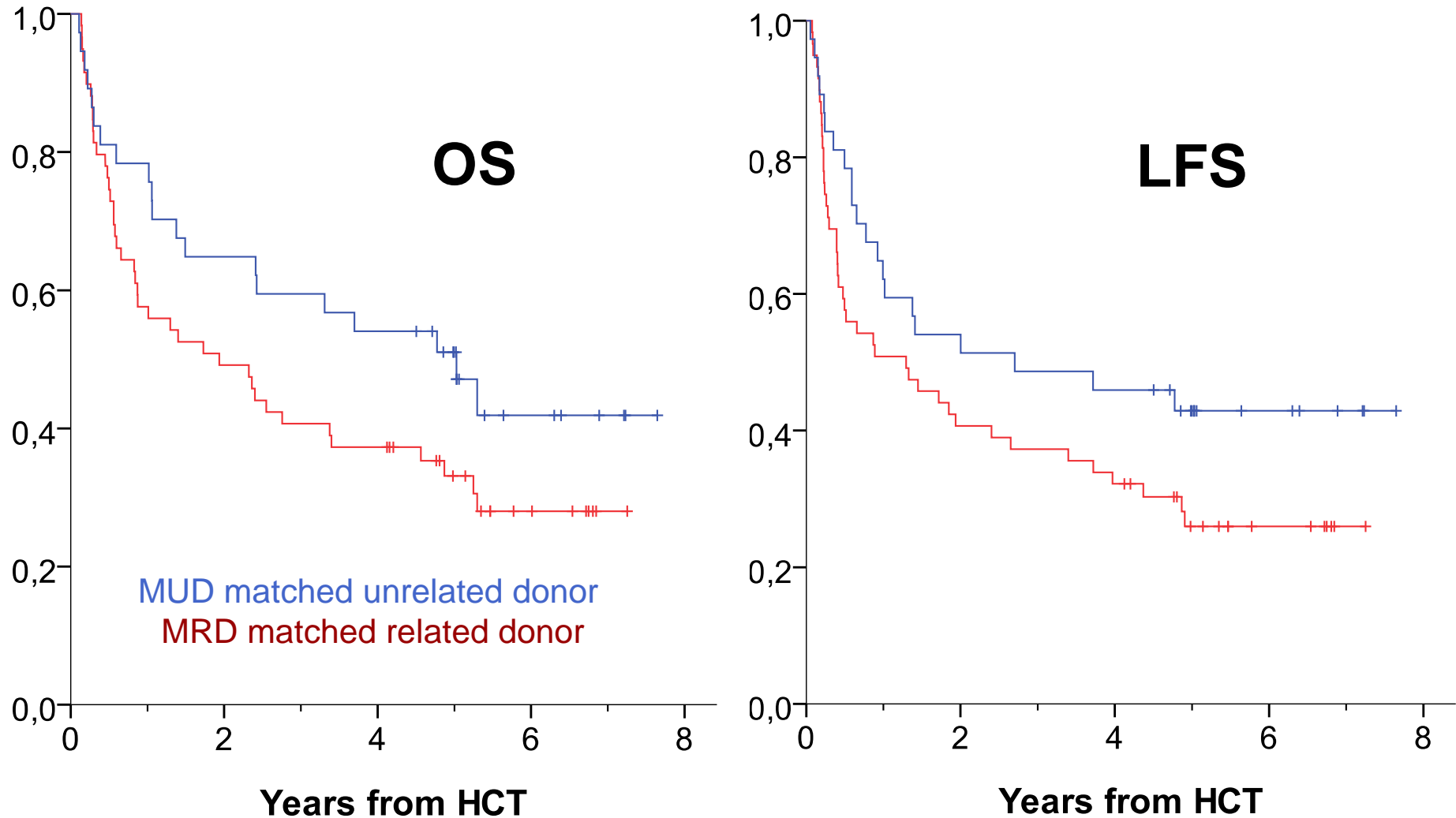
Table 3 Acute and chronic GVHD

Parameter	Acute GVHD, N (%)	Chronic GVHD, N (%)
Incidence	90/167 (53.9)	62/139 (44.6)
Severity		
Grade I	41/90 (45.6)	Limited 28/62 (45.2)
Grade II	15/90 (16.7)	Extensive 34/62 (54.8)
Grade III	27/90 (30)	
Grade IV	7/90 (7.7)	
Organ involved*		
Skin	81/90 (90)	35/62 (56.5)
Stage ≤2	38/81 (47)	
Stage ≥3	23/81 (28.4)	
Liver	18/90 (20)	23/62 (37)
Gut	36/90 (40)	10/62 (16.1)
Others		22/62 (35.5)
Age of patients (years)		
< 50	38/91 (63.7)	37/79 (46.8)
Skin		
Stage ≤2	38/91 (41.8)	
Stage ≥3	13/91 (14.3)	
50-59	15/97 (40.5)	9/31 (29)
Skin		
Stage ≤2	9/97 (24.3)	
Stage ≥3	6/97 (16.2)	
≥60	17/89 (43.6)	16/31 (51.6)
Skin		
Stage ≤2	11/89 (28.2)	
Stage ≥3	4/89 (10.3)	
< 50 vs ≥50	P=0.007	P=0.1
Female donor to male patient (n=34) vs the rest (135)	P=0.9	P=0.2
Age of donors (years) < 50 vs ≥50	P=0.5	P=0.6
NK cell count in the harvest	P=0.95	P=0.8

Abbreviation: NK cells = natural killer cells.  
\*Patients had one or more organ involvement.

# HOVON – OSHO study

Outcome of HCT after TBI based regimens (n=96) in AML



# EBMT study in AML > 60 yrs

If relapse

